# Guideline CarnoCheck® Page 1 of 6 First Time Use of CarnoCheck® V. Grimm Date Rev. 01 Laboratory SOP/equipment/consumables checklist Guideline CarnoCheck® Fage 1 of 6 First Time Use of CarnoCheck® Laboratory SOP/equipment/consumables checklist

# INTRODUCTION

This guideline was written by the manufacturer of CarnoCheck<sup>®</sup> in order to facilitate the setting up of the laboratory equipement prior to the first time use of the CarnoCheck<sup>®</sup> Kit. The compliance with the present guideline should be considered to be a prerequisite for the successful application of CarnoCheck<sup>®</sup>. Furthermore please follow the intructions for use of CarnoCheck<sup>®</sup>.

Part I briefly pictures the separation of the laboratory into four separate rooms whereas Part II describes in detail equipment and consumables necessary within each laboratory room. Part II further specifies additional instructions to be carfully followed. Please proceed with the quideline for installation of the CheckScanner<sup>TM</sup> and the CheckReport<sup>TM</sup> software in order to perform your first analysis with CarnoCheck<sup>®</sup>. Part IV represents a checklist for equipments and consumables.

## **PARTI**

# **ROOM SEPARATION OF THE LABORATORY**

In **Figure 1** the space separation of the laboratory into four separated rooms is highlighted. Every room should be solely utilized for the indicated technique in order to prevent contaminations.

Room 0: SAMPLE COLLECTION AND HOMOGENIZATION

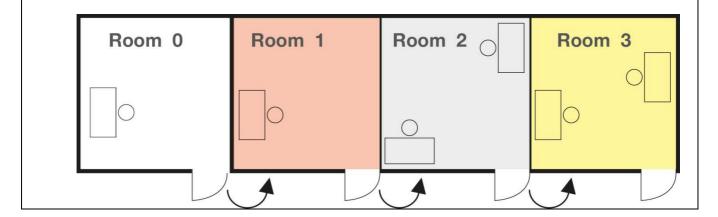
**Room 1: DNA EXTRACTION** 

Room 2: PCR

**Room 3: HYBRIDISATION** 

### Recommendations:

- Usage of a color code for clarification of the separation as well as for avoidance of an interchange of laboratory equipment e.g. micropipettes or reaction tubes
- Change of lab coat after leaving one of the laboratory rooms
- Preferable: integration of a lock between the rooms for homogenisation, DNA extraction and the room for PCR seat up





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PART II								
<b>EQUIPMENT AND CO</b>	NSUN	IABLES						
Room 0: SAMPLE COLLECTION AND HOMOGENIZATION	Sample collection and homogenization has to be performed in this room. After leaving this room the lab coat has to be changed. Owing the fact that within the whole procedur of sample collection and homogenisation is a critical process step in relation to contamination, attention has to be paid to fulfill all instructions delieated in this document. For sample collection we strongly recommend the usage knives with disposable blades. As homogenizer we recommend the usage of laboratory blenders such as the Stomacher® Laboratory homogenizers or other suitable single use equipmen				edure rs			
			precision scales	available available	☐ no			
		0	homogenizer	available	☐ no			
		moo	sample collection devices: knifes	available available	☐ no			
	r and SLES Ro			F and	aluminium foil plastic film bags for homogenizer	available	no	
		EQUIPMENT and CONSUMABLES Room	Lab safety material:	☐ available	□no			
Recommendations for SAMPLE COLLECTION AND HOMOGENIZATION	2	<ol> <li>Use a negative DNA extraction control (water or elution buffer of your extraction kit) which should be treated as a normal sample to verify that your sample batch has not been contaminated during the process.</li> <li>As sample collection devices disposables are recommended.</li> <li>Please carfully follow CarnoCheck® Instructions for Use.</li> </ol>						



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	Standard procedure in room 1:		
Room 1:	The whole DNA extraction procedure with the DNA extraction kit h		
DNA EXTRACTION			
	2.	Parts of the equipment may be replaced by commanufactures. Concerning the heating by	parable instruments of other block, a thermomixer is
		advantageous as it obviats the manual vortexing	
		incubation period. The micropipettes and the	
		should not be used for other purposes than DNA	
	3.	Use either disposable lab coats for DNA extracti	
		leaving this room.	
		M*************************************	
		Micropipette 0.5 -10µl	available no
	_	Micropipette 2 – 20 μl or 0.5 -10μl	available no
		Micropipette 20 – 200 μl	available no
	Soc .	Micropipette 100 – 1000 μl	available no
	Ë	Vortex shaker	available no
		Thermomixer	available no
	EQUIPMENT Room	Microcentrifuge for 1.5 – 2 ml reaction tubes	☐ available ☐ no
	<u>ה</u>	Rack for reaction tubes	available no
	ш	Additional equipment (waste container, timer)	☐ available ☐ no
		DNA Extraction Kit:	☐ available ☐ no
		NucleoSpin Food	
	۲ <b>د</b>	1.5 ml Reaction Tubes	available no
	00	Sterile micropipette filter tips for:	
	<u>~</u>	<ul> <li>Micropipette 2 – 20 μl or 0,5-10 μl</li> <li>Micropipette 100–1000μl</li> </ul>	available no
	CONSUMABLES Room	Ethanol puriss. p.a., ≥ 99,8%	available no
		Lab safety material:	
	Ž	Single use gloves	
	ns:	Single use lab coats	☐ available ☐ no
		Wash bottle with appropriate cleaning	
	Ö	solution for DNA-decontamination	
			_
Recommendations	1 ЛЛ:	ark each part of equipment used in room 1 for DN	IA extraction by a color code
for DNA  in order to prevent accidential equipment exchange		chadach by a dolor dode	
EXTRACTION	2. Us	se only filter tips for pipetting solutions containing i	
		se disposable gloves and change them frequently.	
	4. Pl	ease carfully follow MN Nucleo Spin Food	DNA Extraction Kit and
	Ca	arnoCheck <sup>®</sup> Instructions for Use.	



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Room 2: PCR	In Room 2 the reaction mix for the Polymerase Chain Reaction (PCR) is set bench. The addition of the DNA extracted in room 1 to the PCR reaction out in a separate space within room 2.  After leaving this room the lab coat has to be changed.			to the PCR reaction mix is carried
			Verity <sup>™</sup> 96-Well Thermal Cycler	☐ available ☐ no
			Micropipette 0.5-10 μl (for Taq addition)	available no
			Micropipette 0.2 – 2 μl or 0.5-10 μl (fo addition of DNA eluate)	available no
			Micropipette 5 – 50 μl (division of Mastermix)	available no
			Micropipette 20 – 200 μl (Master-Mix set up)	available no
			Micropipette 100 – 1000 μl	available no
			Clean bench	☐ available ☐ no
		m 2	Vortex shaker	available no
		Rool	Cooling block	available no
		L	Rack for reaction tubes	available no
		ES Room 2 EQUIPMENT Room	Rack for PCR tubes	☐ available ☐ no
			Additional laboratory equipment: waste container	available no
			CarnoCheck <sup>®</sup> Kit	☐ available ☐ no
			AmpliTaq® Gold DNA Polymerase	
			(5U/µI)  1.5 ml Reaction Tubes	available no
Room 2:			Reaction tubes for PCR:	available no
PCR			Single 0.2 ml tubes     8-tube PCR strips	☐ available ☐ no
			Sterile micropipette filter tips for:  • Micropipette 0.5 – 10 µl  • Micropipette 100 – 1000 µl  • Micropipette 5 – 50 µl	☐ available ☐ no
		CONSUMABLES	Lab safety material:	☐ available ☐ no
Recommendations for PCR	<ol> <li>Use only filter tips for pipetting solutions containing DNA.</li> <li>In order to avoid variations due to pipetting errors use pipettes suitable for small volumes. The micropipettes should not be interchanged within the different spaces of Room 2.</li> <li>Vortex and spin down solutions prior to transfering.</li> <li>Please carfully follow CarnoCheck® Instructions for Use.</li> </ol>			



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Room 3: HYBRIDISATION	Within the place. In CheckRep	procedure in room 3:  e third laboratory room the hybridisation reach Room 3 also the CheckScanner oort Software is installed for the final analyse lab coats available.	sis of CarnoCheck <sup>®</sup> . There are
	EQUIPMENT Room 3	Micropipette 2 – 20 μl  Micropipette 10 – 100 μl  Micropipette 100 – 1000 μl  8-channel pipette 5 – 100 μl  Pipettor for glass and plastic pipettes  Rack for PCR tubes  Vortex shaker  Centrifuge for 8-tube PCR strips  Hybridisation Chamber with magnetic slideholder  Temperature controlled water bath for washing step at 50°C  2x oCheck® Washboxes  Handle for Slideholder  Centrifuge for drying the CarnoCheck® chip  Additional laboratory equipment: waste container; timer	□ available         □ no           □ available         □ no
		CheckScanner	available no
		Reaction tubes for hybridisation mix:  Single 0.2 ml tubes 8-tube PCR strips Sterile micropipette filter tips for:	available no
		<ul> <li>Micropipette 2 – 20 μl</li> <li>Micropipette 5 – 100μl</li> </ul>	☐ available ☐ no
	က	Washing of one CarnoCheck® chip: 50 ml Polypropylene Tubes	available no
	Room	Plastic Pipettes for Pipettor to prepare Washing solutions	available no
		50 ml Polypropylene Tubes for drying of the CarnoCheck® chip by centrifugation	available no
	CONSUMABLES	Lab safety material:	☐ available ☐ no
Pacammandations	4 5	lages carfully follow instruction for use	

**HYBRIDIZATION** 

- Please carfully follow instruction for use.
   Use only filter tips for pipetting solutions containing DNA and suitable pipettors.
   Please carfully follow CarnoCheck<sup>®</sup> Instructions for Use.



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# **PART III**

# **ADDITIONAL INSTRUCTIONS**

When implementing currently used state-of-the-art techniques in molecular biology into a laboratory, these instructions must be followed to ensure both maximum safety for laboratory staff and high quality results. These instructions do not substitute the recommendations given in the CarnoCheck® Instruction for Use.

# General Instructions for all rooms:

- Lab coats must be worn throughout the procedures and different sets of lab coats are required for each laboratory room.
- Gloves must be worn during each step of the analysis and must be frequently changed, especially during DNA extraction.
- Lab cleanness: The working place must be decontaminated with an appropriate cleaning solution
- **Reaction tubes:** Never touch the inside of a reaction tube cap.

washing steps.

- Micropipette filter tips have to be sterile.
- Pipetting of small amounts of liquid in the microliter range is a challenge. Therefore take care to pipette with suitable pipettors as accurately as possible. Avoid pipetting less than 2µL.

Labelling: Label every reaction tube in an appropriate manner to ensure traceability.				
Room 1: DNA EXTRACTION	<ul> <li>Carefully follow the instructions for use of the DNA Extraction Kit and CarnoCheck<sup>®</sup>.</li> <li>In addition: <ul> <li>Adjust the temperature of the heating block before the beginning of the DNA extraction procedure.</li> <li>The presence of ethanol may inhibit the PCR reaction afterwards therefore the membrane of the Spin Column has to be dried completely prior to the addition of the final DNA elution buffer.</li> </ul> </li> </ul>			
Room 2: PCR	Carefully follow the protocol for PCR set-up described in detail in CarnoCheck® Instructions for Use.  In addition:  Work under the clean bench while preparing the PCR Reaction Mix			
	<ul> <li>Put on the UV lamp of the clean bench half an hour before you start the PCR set-up.</li> <li>The <i>Taq</i>-Polymerase has always to be stored at -20°C.</li> <li>Keeping the enzyme at room temperature or even on ice could lead to degradation. Therefore, try to keep the time the enzyme is not stored at -20°C as short as possible. Optimally, use a cooling block while pipetting the <i>Taq</i>-Polymerase.</li> <li>Do not use less amount of the <i>Taq</i>-Polymerase than recommended in the CarnoCheck<sup>®</sup> Instructions for Use, otherwise the amplification reaction cannot take place efficiently.</li> <li>Briefly vortex the ingredients for the reaction mix prior and after pipetting.</li> </ul>			
Room 3: HYBRIDISATION	Carefully follow the different steps of hybridisation and washing of the CarnoCheck® chip described in detail in the CarnoCheck® Instructions for Use.  In addition:  Briefly vortex the hybridisation buffer prior and after adding the PCR product.  Airbubbles should be avoided when putting the hybridisation mix on the CarnoCheck® chip.  It is strongly recommended to use an 8-channel multipipette and PCR strips. If more than one CarnoCheck® chip is processed the utilization of an 8-channel multipipette is obligatory.  The surface of the CarnoCheck® chip has to be covered with liquid during all			

The CarnoCheck<sup>®</sup> chip has to be dried completely prior to the analysis with the CheckScanner<sup>TM</sup> and the corresponding CheckReport<sup>TM</sup> Software.